JN THE NAME OF GOD



Faculty of Chemistry Department of Analytical Chemistry

PhD Thesis

Title of the Thesis

Preparation of Nano Quantum Dots, Carbon dots, Protein Nano Fibers, Metal Nano Particles, Carbon Nano Tubes and Organic Compounds Modified Electrodes and their Application to Determination of Pollutants, Medicines and Biological Compounds

> Supervisors: Prof. Mojtaba Shamsipur Prof. Mohammad Bagher Gholivand

> > By: Nader Amini

March 2014

ACKNOWLEDGMENTS

I would like to thank my supervisors Professor Mojtaba Shamsipur and Professor Mohamad Bagher Gholivand for his strong support and encouragement, and for giving me the opportunity to work on very interesting projects. I would like to thank acknowledge the effort of my committee members. I also like to thank Dr Irandoost, Dr Jallali, Professor Farhadi, Professor Fotouhi, Professor Salimi, and Dr Kashaniane for their guidance and cooperation. To all my family I extend my deepest thanks for their constant support. The first of all is my wife for her encouragement, insipiration and love. I am grateful to all my friends, M.Sc and Ph.D students in analytical chemistry research laboratory. Also, I would like to express my special thanks to, Mr Abdollah Khatoni, Dr Nassri, Dr Roushani, Dr Sageghi, Dr. Rajabi, Dr Pashabadi, Dr Mohammadi, Dr Habibi, Dr Barati, Dr Mandomi, Dr Dehdashtian, Dr Hashemi and Dr Hallaj for their friendship and support.

I extend thanks to Miss chamani; finally the author wishes to express his gratitude to department of chemistry of Razi University.

Nader Amini

Dedicate to the most important persons in my life:

My Wife & My Son (KAREN Kocholou)

Abstract

Glycation induced bovine serum albumin in which fibrilogenesis (nano fibrils) followed by fluorescence (Thioflavin T) and also by using dynamic light scattering (DLS) and transmission electron microscopy (TEM) to achieve the size and morphology of fibrils, respectively. A novel electrochemical biosensor for the detection of hydrogen peroxide was proposed based on immobilizing poly (alizarin yellow R)/nano-fibris on glassy carbon electrode. Cyclic voltammetry (CV) and amperommetry were used to confirm the successful stepwise assembly procedure of the biosensor. The electrocatalytical behaviors of the sensor were also investigated by cyclic voltammetry and amperommetry. Results showed that poly (alizarin yellow R)/ nano-fibs exhibited a remarkable electrocatalytic activity for the reduction of hydrogen peroxide under optimal conditions. The electrocatalytic response of the sensor was proportional to the hydrogen peroxide concentration in the range of (1 μ M to 2.2 mM) with a limit of detection and sensitivity of 0.29 μ M and 0.024 μ A/ μ M, respectively. The modified electrode showed many advantages such as simple preparation, high sensitivity, low detection of limit, excellent catalytic activity at physiological pH values and short response time.

A novel electrochemical sensor for the detection of hydrazine was proposed based on immobilizing ZnS/Mn quantum dots and multi wall carbon nanotube (MWCNT) on glassy carbon (GC) electrode. Scanning electron microscopy (SEM), Transmission electron microscopy (TEM), electrochemical impedance spectroscopy (EIS) , cyclic voltammetry (CV) were used to confirm the successful stepwise assembly procedure of the sensor .The electrocatalytic behaviors of the sensor was also investigated by cyclic voltammetry and differential puls voltammetry .Tests showed that hydrazine by (zinc sulfide doped with manganese) Quantum Dots /multi wall carbon nanotube (ZnS/Mn QDs-MWCNT) exhibited a remarkable electrocatalytic activity for the oxidation of hydrazine. Under optimal conditions, the electrocatalytic response of the sensor was proportional to the hydrazine concentration in the range of 0.09 to 1.2μ M. With a detection limit and sensitivity of 28nM and 0.0009μ A μ M⁻¹. This electrode shows many advantages such as simple preparation, high sensitivity, excellent catalytic activity at pH 7 and antifouling property toward hydrazine and its oxidation product.

A novel electrochemical sensor for the detection of l-cysteine was proposed based on immobilizing poly (alizarin yellow R)/carbon quantum dots on glassy carbon electrode. Hydrothermal treatment was used to prepare carbon quantum dots (CQDs). Transmission electron microscopy (TEM) and FTIR were used for characterization of carbon quantum

dots. Electrochemical impedance spectroscopy, cyclic voltammetry (CV) and amperommetry were used to confirm the successful stepwise assembly procedure of the sensor. The electrocatalytic behaviors of the sensor were also investigated by cyclic voltammetry and amperommetry. Results showed that poly (alizarin yellow R)/carbon dots exhibited a remarkable electrocatalytic activity for the oxidation of 1-cysteine under optimal conditions. The electrocatalytic response of the sensor was proportional to the 1-cysteine concentration in the range of (0.3 to 3.6μ M) and (3.9 to 7.2μ M) with a limit of detection and sensitivity of 90 nM and 0.482μ A/ μ M, respectively. The modified electrode show many advantages such as simple preparation, high sensitivity, low detection of limit, excellent catalytic activity at physiological pH values, short response time, and remarkable antifouling property toward 1-cysteine and its oxidation product.

For the first time, a nonenzymatic electrochemical sensor for the detection of lysine was proposed based on immobilizing Multi wall carbon nanotube (MWCNT) and Titanium oxide nanoparticles (TiO₂NPs) on glassy carbon (GC) electrode. Scaning electron microscopy (SEM) and electrochemical impedance spectroscopy (EIS) were used to confirm the successful stepwise assembly procedure of the sensor. The electrocatalytical behaviors of the sensor were also investigated by cyclic voltammetry (CV) and differential pulse voltammetry (DPV). The results showed that MWCNT- TiO₂NPs exhibited a remarkable electrocatalytic activity for the oxidation of lysine. Under optimal conditions, the DPV response of the sensor was proportional to the lysine concentration in the range of 500 to 5500 nanomolar with a detection limit and sensitivity of 390 nM and $0.1795\mu A\mu M^{-1}$. This electrode show many advantages such as simple preparation without using any enzyme special electron transfer mediator or specific reagent, excellent catalytic activity at physiological pH values and antifouling property toward lysine and its oxidation product. Furthermore, the selectivity of the proposed sensor was tested in the presence of some amino acids.

A novel electrochemical sensor for the detection of insulin was proposed based on immobilizing silica nanoparticles/Nafion on glassy carbon electrode. Transmission electron microscopy, electrochemical impedance spectroscopy, cyclic voltammetry (CV) and differential pulse voltammetry (DPV) were used to confirm the successful stepwise assembly procedure of the sensor. The electrocatalytical behaviors of the sensor were also investigated by CV and DPV. Results showed that nano-SiO₂ exhibited a remarkable electrocatalytic activity for the oxidation of insulin under optimal conditions. The electrocatalytic response of the sensor was proportional to the insulin concentration in the

range of 10 to 50 nM with a limit of detection and sensitivity of 3.1 nM and 300 pAnM⁻¹, respectively. The modified electrode show many advantages such as simple preparation without using any special electron transfer mediator or specific reagent, high sensitivity, excellent catalytic activity at physiological pH values, short response time, and remarkable antifouling property toward insulin and its oxidation product.

The electrochemical behavior of chloropromazine at glassy carbon (GC) electrode nanoparticles/ chloropromazine/ modified with silica Nafion (SNPs/CPZ/Nf) nanocomposite was investigated. The apparent electron transfer rate constant (ks), transfer coefficient (α) and surface concentration (Γ c) were found to be 0.56 s⁻¹, 0.49 and 3.49 × 10^{-7} molcm²⁻, respectively. Cyclic voltammetry technique has been used for stabilization of nanocomposite on the surface GC electrode. Transmission electron microscopy (TEM), electrochemical impedance spectroscopy (EIS), cyclic voltammetry (CV) and differential pulse voltammetry techniques were used to confirm the successful stepwise assembly procedure of the electrode. The modified electrode showed electrocatalytic activity toward nitrite electro-reduction at 0.12V. The detection limit (signal to noise) and sensitivity are 7μ M and 0.0007μ A/ μ M, respectively. The advantages of the nitrite amperometric detector based on the SNPs/CPZ/Nf nanocomposite GCE are a low detection limit, especially a reduction in low potential, high sensitivity and inherent stability at pH 2, catalytic activity for nitrite reduction antifouling property toward nitrite and its reduction product. Furthermore, the proposed electrode was used for determination of nitrite in food samples.

The electrochemical behavior of chloropromazine as a modifier on the surface of electrode was investigated. The electrochemical properties of chloropromazine in to the silica nanoparticles/ chloropromazine/ nafion (SNPs/CPZ/Nf) nanocomposite at pH 2-10 were investigated at a glassy carbon electrode. Well defined reversible redox couples were observed in acidic solutions and irreversible in alkaline solutions. The (SNPs/CPZ/Nf) nanocomposite modified electrodes were characterized with a transmission electron microscopy (TEM), electrochemical impedance spectroscopy (EIS) and cyclic voltammetry (CV). The apparent electron transfer rate constant (k_s), transfer coefficient (α) and the surface concentration (Γ c) were determined by cyclic voltammetry and they were about 0.025 s⁻¹, 0.50 and 1.26 × 10⁻⁶ molcm²⁻, respectively. Moreover, electrocatalytic oxidation of sulfide on the surface of modified electrode was investigated with cyclic voltammetry and amperometry methods at pH=7. The detection limit (signal to noise) and sensitivity are 90nM and 0.0021nA/ μ M, respectively. The prepared modified electrode

showed several advantages, such as a simple preparation method, high sensitivity, very low detection limits and excellent reproducibility. Moreover, the proposed sensor can be used for sulfide analysis in water samples.

Table of contents

Title	Page
Chapter 1: INTRODUCTION & LITERATURE REVIEW	
PART A: Theory and Analytical Applications of Modified Electrodes	
1. The Theory of Mediated Electrocatalysis at Modified Electrodes	3
PART: B: Chemically and Physically Modified Electrodes	
1.2. Introduction	14
1.3. Electrocatalytic aspects	15
1.4. Characterization	16
1.4.1. Electrochemical methods	16
1.4.2. Spectrochemical and other methods	17
1.5. Physical modification	18
1.5.1. Physical surface modification	18
1.5.2. Physical environment modification	18
1.6. Chemical modification	19
1.6.1. Principal routes	20
1.6.2. Metal (hydr) oxides	22
1.6.3. Carbons (glassy carbon, (pyrolytic) graphite)	23
1.6.4. Polymer modification	24
1.6.4.1. Preparation	24
1.6.4.2. Mechanism of polymerization	25
1.6.4.3. Mechanism of charging	25
1.6.5. Conductivity	26
1.6.5.1. Electrocatalytic activity	26
1.7. Built-in electrocatalysts	28
1.8. Concentration profiles in modified electrodes	29
1.9. Some applications	
1.9.1. Analysis	29
1.9.1.1. Salt bridge-free reference electrode for use in, qon-aqu	eous solvents
[51]	29
1.9.1.2. Glucose sensor in blood	
1.9.2. Synthesis	
1.9.2.1. Synthesis of hydrogen peroxide	

1.9.2.2. Stereo- and enantio-selective synthesis	31
1.9.3. Bio- and photo-electrochemistry	
1.10. Concluding remarks	32
PART C: Application of Nanoparticles in Electrochemical Sensors and	l Biosensors
1.11. Introduction	
1.12. The Functions of Nanoparticles	35
1.12.1 Immobilization of Biomolecules	
1.12. 2. Catalysis of Electrochemical Reactions	
1.12. 3. Enhancement of Electron Transfer	40
1.12.4. Labeling Biomolecules	42
1.12. 5. Nanoparticles Acting as Reactant	45

Chapter 2: Electropolymerization of Alizarin Yellow on Glycation Induced Nanofibrils for Preparation of a Novel Hydrogen Peroxide Biosensor

2. Introduction
2.1. Experimental
2.1.1. Reagents
2.1. 2. Apparatus
2.1.3. Fibril preparation
2.2. Preparation of the modified electrode
2.2.1. Electropolymerization of the alizarin yellow R on nano-fibs/ GCE52
2.3. Results and discussion
2.3.1. Characterization of Nano-fibrils
2.3.2. Fluorescence analysis
2.3.3. Transmission Electron Microscopy (TEM)54
2.4. Electrochemical characterization of poly (alizarin yellow R)/nano-fibs /GCE55
2.5. Electrocatalytic oxidation of HP on poly (alizarin yellow R)/nano-fibs /GCE57
2.6. Amperometric detection of hydrogen peroxide at poly (alizarin yellow R)/ nano-
fibs /GCE61
2.7. Comparison of the figures of merit of the proposed sensor with those of previous
electrochemical methods
2.8. Determination of HP in real samples

Chapter 3: Nanomolar Detection of Hydrazine by (Zinc Sulfide Doped with Manganese) Quantum Dots /Multi Wall Carbon Nanotube Composite on Glassy Carbon Electrode

3. Introduction	66
3.1. Experimental	67
3.1.1. Reagents	67
3.1.2 Apparatus	68
3.1.3. Synthesis of ZnS/Mn quantum dots	68
3.1.4. Preparation of the modified electrode	68
3.2. Results and discussion	69
3.2.1. Characterization of ZnS/Mn quantum dots	69
3.2.2. Morphological and electrochemical characterization of ZnS/Mn	QDs -
MWCNTs /GC electrode	69
3.2.3. Electrocatalytic oxidation of hydrazine on ZnS/Mn QDs -MWCN	Ts GC
electrode	71
3.2.4. Repeatability and stability	76
3.3. Interference effects	76
3.4. Comparison of the figures of merit of the proposed sensor with those of p	revious
modified electrodes	77
3.5. Application	77
3.6. Conclusion	78

Chapter 4: Nanomolar Detection of L-cysteine Using a Glassy Carbon Electrode Modified with Carbon Quantum Dots and Poly Alizarin Yellow R

4. Introduction	80
4.1 Experimental	82
4.1.1. Reagents	82
4.1.2. Apparatus	82
4.1.3. Synthesis of carbon quantum dots	82
4.1.4. Preparation of modified electrode	82
4.2. Results and discussion	83
4.2.1 Characterization of carbon quantum dots	83
4.2.2. Electrochemical characterization of poly (alizarin yellow	R)/carbon dots
/GCE	84

4.2.3. Electrocatalytic oxidation of cysteine on poly (alizarin yellow R)/carbon
dots /GCE
4.2.4. Amperometric detection of cysteine at poly (alizarin yellow R)/carbon dots
/GCE
4.3. Repeatability and stability
4.4. Comparison of the figures of merit of the proposed sensor with those of previous
electrochemical methods
4.5. Analytical application
4.6. Conclusion

Chapter 5: Nonenzymatic L-lysine Amino Acid Detection Using Titanium Oxide Nanoparticles/ Mmulti Wall Carbon Nanotube Composite Electrodes

5. Introduction	94
5.1. Experimental	
5.1.1. Reagents	95
5.1.2. Apparatus	96
5.1.3. Preparation of the GC modified electrode	96
5.2. Results and discussion	96
5.2.1. Morphological and electrochemical characterization of MW	CNT- TiO ₂ NPs
/GC electrode	96
5.2.2. Electrocatalytic oxidation of L-lysine on MWCNT-	TiO ₂ NPs /GC
electrode	
5.2.3. Determination of Lys by differential pulse voltammetry	104
5.3. Interference effects	
5.4. Repeatability and stability	105
5.5 Comparison of the figures of merit of the proposed sensor with th	ose of previous
enzymatic methods	106
5.6- Analytical applications	106
5.7. Conclusion	107

Chapter 6: Electrocatalytic Determination of Traces of Insulin Using a Novel Silica Nanoparticles-Nafion Modified Glassy Carbon Electrode

6.	Introduction	109
6	5.2. Experimental	111

6.2.1. Reagents	111
6.2.2. Apparatus	111
6.2.3. Preparation of the modified electrode	111
6.3. Results and discussion	112
6.3.1. Morphological and electrochemical characterization of	SiO ₂ NPs-
Nafion/GCE	112
6.3.2. Electrocatalytic oxidation of insulin on SiO ₂ NPs-Nafion/GCE	113
6.4. Interference study	118
6.5. Repeatability and stability	118
6.6. Comparison of the figures of merit of the proposed sensor with those of	of previous
electrochemical methods	119
6.7. Analytical applications	119
6.8. Conclusion	121

Chapter 7: Electrochemical Properties of Chlorpromazine in Silica Nanoparticles/Chloropromazine/Nafion Nanocomposite: Application to Nitrite Detection

7. Introduction
7.1.Experimental
7.1.1. Reagents
7.1.2. Apparatus
7.1.3. Preparation of the modified electrode125
7.2. Results and discussion
7.2.1. Morphological and electrochemical characterization of SNPs/CPZ/Nf/GC
electrode126
7.2.2. Electrochemical behavior of the SNPs/CPZ/Nf nanocomposite modified
GC electrode
7.2.3. Stability and pH dependence of the SNPs/CPZ/Nf nanocomposite modified
GC electrode
7.3. Electrocatalytic reduction of nitrite on SNPs/CPZ/Nf nanocomposite modified
GC electrode
7.4. Interference effects
7.5. Comparison of the figures of merit of the proposed sensor with those of previous
modified electrodes

7.6. Application in food Analysis	139
7.7. Conclusion	140

Chapter 8: Electrocatalytic Oxidation of Sulfide and Electrochemical Behavior of
Chloropromazine Based on Organic–inorganic Hybrid Nanocomposit
8. Introduction
8.1. Experimental144
8.1.1. Reagents
8.1.2. Apparatus
8.1.3. Preparation of the modified electrode145
8.2. Results and discussion145
8.2.1. Morphological and electrochemical characterization of SNPs/CPZ/Nf/GC
electrode145
8.2.2. Electrochemical behavior of the SNPs/CPZ/Nf nanocomposite modified
GC electrode
8.2.3. Stability and pH dependence of the SNPs/CPZ/Nf nanocomposite
modified GC electrode
8.3. Electrocatalytic oxidation of sulfide on SNPs/CPZ/Nf nanocomposite modified
GC electrode152
8.3.1. Amperometric response to sulfide155
8.4. Comparison of the figures of merit of the proposed sensor with those of previous
electrochemical methods156
8.5. Interference effects157
8.6. Repeatability and lifetime157
8.7. Application158
8.8. Conclusion
References

List of Figures

Figures Page
Fig.1. 1. Schematic representation of the processes possible at redox polymer modified
electrodes [10]
Fig.1.2. Locations o f the possible reaction zones at redox polymer modified electrodes and
the notation used to describe them
Fig.1.3Kinetic zone diagram showing the kinetic control of the processes taking place in
the redox polymer ilm as a function of surface coverage, Γ9
Fig.1.4.Diagnosis of mechanism for redox polymer modified electrodes11
Fig. 1.5. Cyclovoltammograms for several types of platinum in $1 \text{ M H}_2\text{SO}_4$ with a potential
scan rate of 10mVs ⁻¹ . Continuous scanning results in a surface lattice structure with
predominant Pt(100); own results17
Fig.1.6. Cathodic reduction ofdioxygen[37]21
Fig1.7. Chemical modification with vinyl or amine compounds at a highly, by rf-Ar
plasma, activated carbon surface. R is a functional group
Fig.1.8, Formation of a monolayer (a) or an oligomeric structure (b), making use of
organosilanes as bridge molecules
Fig.1.9. Functionalities at the edge plane of pyrolytic graphite
Fig.1.10. Making a peptide or an ester bond by using sulphonyl chloride and/or the
dehydrating agent DCC (dicyclohexylcarbodiimide)23
Fig. 1.11. Attaching of the bridge molecule cyanuric chloride at a pretreated carbon
surface
Fig.1.12. Cyclovoltammogram of a $0.2\mu m$ PPy-layer on platinm in $0.1M$ Bu ₄ N
BF_4 /acetonitrile; v = 20mVs ⁻¹ 25
Fig.1.13. Redox polymer electrode with protonated poly(4-vinylpyridine) as polymer
matrix and $X^{-} = \text{CoTSPc}^{4-}$ as counter-ion/ redox centre
Fig.1.14. Immobilized Ru(edta)(OH ₂) redox complex at a pyrolytic graphite (Cp)
electrode, modified with poly(4-vinylpyridine)27
Fig.1.15. Concentration profiles for S (-) and C(R) ()
Fig.1.16. (a) Modified glucose oxidase; (b) sensor
Fig.1.17. Modified electrode for the reduction of dioxygen to hydrogenperoxide (silane
bridged naphthoquinone)

Fig.1.18. Preferential p-substitution of chlorine in anisole at a cyclodextrine modified Fig.1.19. Photo-electrolytic splitting of water with semiconducting particles partly covered Fig.1.20.The immobilization of DNA with gold nanoparticles (adapted from [22] with Fig.1.21. Electrical coupling of gold nanoparticle-reconstituted glucose oxidase to an electrode by a) the adsorption of gold nanoparticle-reconstituted glucose oxidase to a dithiol monolayer associated with the gold electrode and b) the adsorption of gold nanoparticles functionalized with FAD on the dithiol-modified gold electrode followed by Fig.1.22. Procedure of a noncompetitive heterogeneous electrochemical immunoassay Fig.2.1. The formation of amyloid fibrils. Anatively folded monomer undergoes a conformational transition into a β -sheet-rich state, usually through a partial unfolded state. Self-assembly of these intermediates into ladders of β strands results in the formation of Fig.2.2. The continuous CVs for the electropolymerization of alizarin yellow R on nanofibs/ GCE in 0.1 M phosphate buffered (pH11) at 100 mVs⁻¹......55 Fig. 2.4. TEM image of nanofibrils......55 Fig.2.6. Electrochemical impedance spectroscopy (EIS) of (a) bare GC electrode (b)nano-Fig.2.7. Cyclic voltmmetry response of unmodified GCE in the absence of HP (a) unmodified (b) polyalizarine yellow R modified (c) nano-fibs modified (d) and (e) polyalizarin yellow / nano-fibs modified GC electrode in the presence 2.3mM of HP, scan Fig.2.9. Cyclic voltammogram of alizarin / nano-fibs/ GC modified electrode in pH 6 buffer containing different concentration of HP from 50 to 2100 µM. inset, plot of peak

Fig.2.10. (a) Cyclic voltammetry response of a GCE modified with alizarin / nano-fibriles modified GC in a phosphate buffer (pH 6) containing 500µM of HP at different scan rates Fig.2.11.Amperometric response at rotating poly (alizarin yellow R)/ nano-fibriles modified GC electrode (rotation speed 2000 rpm) held at -0.45 V in buffer solution (pH6) for successive addition of (A) 150µM HP (B) 1µM HP (a and b), Plot of Fig.3.1.(A) UV-vis absorption (a), fluorescence emission (b) and room temperature phosphorescence (c) spectra of L-cysteine capped Mn-doped ZnS QDs. (B) TEM image of L-cysteine capped Mn-doped ZnS QDs......69 Fig.3.2. The SEM of MWCNT (a) Mn-doped ZnS QDs / MWCNT (b) respectively......70 Fig.3.3. Electrochemical impedance spectroscopy (EIS) of (a) bare GC electrode (b) Mn-Fig.3.4. Cyclic voltammogram of (a) bare (b) MWCNT (c) Mn-doped ZnS QDs / MWCNT GC electrode in pH = 7 buffer containing 170 μ M of hydrazine......72 Fig.3.5. The plot of a GCE modified in 0.1 M phosphate buffer solution for values pH (2, 3, 4, 5, 6, 7, 8 and 9).....73 Fig.3.6. Cyclic voltammogram of Mn-doped ZnS QDs / MWCNT modified GC electrode in pH = 7 buffer containing different concentration of hydrazine from 10 and 180 μ M. Fig.3.7. (a) Cyclic voltammetry response of a GCE modified with Mn-doped ZnS QDs / MWCNT in a phosphate buffer (pH 7) containing 100µM of hydrazine at different scan Fig.3.8. DPV of Mn-doped ZnS QDs / MWCNT modified GC electrode in pH 7 buffer containing different concentration of hydrazine from 100 to 1200 nM. Inset, plot of peak Fig.4.1. TEM image of carbon QDs......83 Fig.4.3. The continuous CVs for the electropolymerization of alizarin yellow R on carbon dots GCE in 0.1 M phosphate buffere (pH 11) at potential scan rate 100 mVs⁻¹......85 Fig.4.4 Electrochemical impedance spectroscopy (EIS) of (a) bare GC electrode (b) carbon Fig.4.5. Cyclic voltammograms of unmodified electrode in the absence of cysteine (a) unmodified (b) carbon QDs modified (c) alizarin yellow modified (d) and (e) carbon QDs /

alizarin yellow modified GC electrode in the presence 50 μ M of cysteine , scan rate 20 Fig.4.6. (a) Cyclic voltammetry response of a GCE modified with carbon QDs / alizarin in a phosphate buffer (pH 7) containing 50 µM of cysteine at different scan rates (from inner Fig.4.7. Cyclic voltammogram of carbon QDs / alizarin modified GC electrode in pH = 7buffer containing different concentration of cysteine from 0.6 to 12 µM. inset, plot of peak Fig.4.8. Amperommetric responses at rotating modified GC electrode (rotating speed 2000rpm) held at 0.55V in buffer solution (pH 7). Inset, Plot of the peak currents as a Fig. 5.1. SEM images of (a) MWCNT/GCE; (b) MWCNT- TiO₂NPs /GCE......97 Fig.5.2. Electrochemical impedance spectroscopy responses of bare GCE (a) and GCE/MWCNT (b) and MWCNT- TiO₂NPs /GCE in 5 mM [Fe(CN)₆]^{3-/4-} redox couple. Fig.5.3. Cyclic voltammetry response of unmodified electrode (a) MWCNT modified (b) and MWCNT- TiO₂NPs modified(c) GC electrode in the presence 110 µM of lysine, scan Fig.5.4. Cyclic voltammograms of MWCNT- TiO2NPs /GC electrode in different pH solutions, from left to right, 2 to 11, in the presence of 40 μ M of lysine and scan rate of 20 mVs^{-1} .(a), plot of peak current vs. pH values(b), plot of potential vs. pH values.....101 Fig.5.5. Cyclic voltammogram of MWCNT- TiO₂NPs /GC electrode in pH = 7 buffer containing different concentration of lysine from 5 to 40 µM and 40 to 110µM. Inset, plot Fig.5.6. (a) Cyclic voltammetry response of a GCE modified with MWCNT- TiO₂NPs in a phosphate buffer (pH 7) containing 50µM of lysine at different scan rates (from inner to outer) 10-100 mVs⁻¹. Inset, (A) plot of peak current vs, $v^{1/2}$. (B) Ep vs, Logv.....104 Fig5.7 .DPV of $TiO_2/MWCNT$ modified GC electrode in pH = 7 buffer containing different concentration of lysine from 0.5, 1, 1.5, 2, 2.5 to 5.5 µM. inset, plot of peak Fig. 6. 1. TEM image of SiO₂NPs-Nafion/GCE......112 Fig. 6.2. Electrochemical impedance spectroscopy responses of bare GCE (a) and SiO₂NPs-Nafion GCE......113

Fig. 6.3. Cyclic voltammetry responses of the bare GCE (a and b) and modified SiO ₂ NPs-
Nafion/GCE electrodes (c and d) in the absence (a and c) and presence 50 μ M of insulin (b
and d) at a scan rate of 30 mV s ⁻¹ and a pH of 7.35114
Fig. 6,4. The CVs of a 50 μ M solution of insulin at different pH values (from 7 to 9) at the
SiO2NPs modified GC electrode
Fig.6. 5. Cyclic voltammograms of SiO2NPs-Nafion/GCE in a 0.1 M buffer solutions of
pH 7.35 containing varying concentrations of insulin: (a) 0.0, (b) 0.1,(c) 0.2, (d) 0.3, (e)
0.4, (f) 0.5, (g) 0.6, (h) 0.7, (i) 0.8 μ M. Inset shows a linear plot of peak current vs. insulin
concentration116
Fig. 6.6. Cyclic voltammetry responses SiO2NPs-Nafion/GCE in a 0.1 M phosphate buffer
solution of pH 7.35 containing 0.1µM of insulin at different scan rates: (a) 10, (b) 20,(c)
30, (d) 40, (e) 50, (f) 60, (g) 70, (h) 80, (i) 90, (j) 100 mV s-1. Inset shows a linear plot of
peak current vs. $v^{1/2}$
Fig. 6.7. Differential pulse voltammograms SiO ₂ NPs-Nafion/GCE in a 0.1 M phosphate
buffer solution of pH 7.35 containing different concentration of insulin: 0, 10, 20, 30, 40,
50 nM. Inset shows a linear plot of peak current vs. insulin concentration118
Fig.7.1. The TEM of SNPs (a) SNPs/CPZ/Nf nanocomposite (b) respectively126
Fig.7.2. Electrochemical impedance spectroscopy(EIS) of (a) bare GC electrode (b)
Nf/GC electrode; (c) SNPS/Nf/GC electrode SNPS/CPZ/Nf/GC electrode(d) in 5 mM
probe $Fe(CN)_6^{4-/3-}$ (Inset)Fitted circuit for the electrochemical impedance spectroscopic
data127
Fig. 7.3. Consecutive cyclic voltammograms of a glassy carbon electrode modified with
SNPs/CPZ/Nf/GCE (a) GCE bare and SNPs/Nf/GCE (b) in phosphate buffer solution
(pH 2), scan rate100 mVs ⁻¹ 129
Fig.7.4. (a) Cyclic voltammetry response of a GCE modified with SNPs/CPZ/Nf
nanocomposite in a 0.15 M phosphate buffer (pH 2) at scan rates of (inner to outer) 10, 20,
30, 40, 50, 60, 70, 80, 90, 100mVs^{-1} .(b) and (c) plots of peak current vs. scan rate and
square root of scan rate
Fig.7.5. The variation of $\Delta E_p(E_p\text{-}E^\circ)$ vs . logv for the modified GCE. Inset is the same
plote at higher sweep rates
Fig.7.6. Cyclic voltammograms of a GCE modified in 0.15 M phosphate buffer solution
for values pH (2, 3, 4, 5, 6, 7, 8, and 9). inset; plot of E° vs pH132

Fig.7.7. Cyclic voltammograms of SNPs/CPZ/Nf nanocomposite modified GCE in buffer solution pH 2 at scan rate of 20 mVs⁻¹ in the absence(c) and presence of 500µM nitrite(d), Fig.7.8. Cyclic voltammograms of SNPs/CPZ/Nf nanocomposite modified GCE in buffer solution pH 2 at scan rate of 20mVs⁻¹ with increasing nitrite concentration (from inner to outer) 0.0, 100, 200, 300, 400, 500,600 and 700 µM. inset; plot of peak current vs. nitrite Fig.7.9. Peak current of a GCE modified in 0.15 M phosphate buffer solution and 500 µM nitrite for values pH (2, 3, 4, 5, 6, 7 and 8).....136 Fig.7.10. (a) cyclic voltammetry response of a GCE modified with SNPs/CPZ/Nf nanocomposite in a 0.15 M phosphate buffer (pH 2) containing 500 µM of nitrite at different scan rates (from inner to outer) 10-100 mVs⁻¹. (b) Plot of I_p vs. $v^{1/2}$. (c) Plot of E_p Fig.7.11. DPV of SNPs/CPZ/Nf nanocomposite modified GC electrode in pH 2 buffer containing different concentration of nitrite 0.0, 20, 30, 40, 50 and 60 μ M. Inset, plot of Fig.8.1. The TEM of SNPs (a) SNPs/CPZ/Nf nanocomposite (b) respectively......146 Fig.8.2. Electrochemical impedance spectroscopy(EIS) of (a) equal circuit (b) bare GC Fig. 8.3. (a) Cyclic voltammetry response of a GCE modified with SNPs/CPZ/Nf nanocomposite in a phosphate buffer (pH2) at scan rates of (inner to outer) 10, 20, 30, 40, 50, 60, 70, 80, 90, 100 mVs⁻¹.(b) plots of peak current vs. square root of scan rate......149 Fig.8.4. The variation of $\Delta E_n(E_n - E^\circ)$ vs. logv for the modified GCE. Inset is the same plots Fig.8.5. Cyclic voltammograms of a GCE modified in phosphate buffer solution for values pH (2, 3, 4, 5, 6, 7, 8, 9 and 10).....151 Fig.8.6. Cyclic voltammograms of SNPs/CPZ/Nf nanocomposite modified GCE in buffer solution pH 7 at scan rate of 10mVs^{-1} in the absence(c) and presence of 6 μ M sulfide(d), (a and b) are same results as (c and d) for bare GC electrode......152 Fig.8.7. Peak current of a GCE modified in phosphate buffer solution and 6 µM sulfide for values pH (2, 3, 4, 5, 6, 7, 8 and 9).....153 Fig.8.8. Cyclic voltammograms of SNPs/CPZ/Nf nanocomposite modified GCE in buffer solution pH 7 at scan rate of 10mVs⁻¹ with increasing sulfide concentration (from inner to outer) 0.0, 1, 2, 3, 4, 5 and 6 µM. inset; plot of peak current vs. sulfide concentrations..154